

UNITED STATES DEPARTMENT OF AGRICULTURE
FOOD SAFETY AND INSPECTION SERVICE
WASHINGTON, DC

<h1>FSIS NOTICE</h1>	16-12	2-29-12
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**NATIONWIDE RAW LIQUID EGG MICROBIOLOGICAL BASELINE SURVEY –
UPDATE**

NOTE: DO NOT IMPLEMENT THIS NOTICE UNTIL MARCH 12, 2012

I. PURPOSE

This notice announces that the Food Safety and Inspection Service's (FSIS) Nationwide Raw Liquid Egg Microbiological Baseline Survey (RLEBS) will begin on March 12, 2012, and that the 90-day training ("shake down") period referred to in FSIS Notice 52-11, nationwide FSIS RLEBS – Shakedown, has ended.

NOTE: This notice applies only to inspection program personnel (IPP) at plants that are included in the nationwide FSIS RLEBS. Only plants that pasteurize or heat treat dried egg products are included.

KEY POINTS

- *FSIS Notice 52-11 is cancelled.*
- *Only egg products plants that pasteurize or heat treat dried egg products, will be included in the baseline survey. Egg products plants that only break shell eggs will not be included in the baseline survey.*
- *The sample container to be used has been changed to ensure a more secure seal and prevent leakage during sample shipment.*
- *Samples are to be collected on both shifts in multiple shift establishments.*
- *Each sample will consist of one 400 ml sample. The sample is to be collected prior to the addition of any ingredients and prior to pasteurization or heat treatment, which may require the sample to be collected the day before pasteurization.*
- *When the lot of egg product to be sampled is to be dried, the sample is to be*

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OPI: OPPD

collected prior to the start of any further processing of the liquid egg (e.g., desugaring, reverse osmosis).

- *Samples may be collected on the day prior to pasteurization when the plant's production practices or further processing procedures are such that a sample cannot be collected on the same day that the product is to be pasteurized or heat treated.*
- *Both domestically broken raw egg products and imported raw egg products will be sampled during this survey.*
- *FSIS Form 10,210-3, Sample Request Form, contains survey questions to be answered by IPP collecting the sample.*
- *Survey sample results are not posted in the Laboratory Electronic Application for Results Notification (LEARN).*
- *Survey samples are not used for regulatory purposes.*
- *All supply requests and questions are to be submitted to the "Liquid Egg Baseline" mailbox in Outlook.*
- *IPP are to continue to collect samples for the RLEBS in addition to other regulatory egg product samples.*

II. BACKGROUND

A. During the baseline survey, IPP are to collect samples of unpasteurized liquid egg product (i.e., liquid whole egg, liquid egg white, or liquid egg yolk) from shell eggs broken in official egg product plants and imported products received before ingredients are added and prior to pasteurization or heat treatment. The survey will last for a minimum of 12 months. Pasteurization or heat treatment is the processing step that subjects egg products to elevated temperatures to reduce *Salmonella* spp. to a level that is not likely to present a public health risk under normal conditions of distribution and storage. The pasteurization or heat treatment of egg products can be done in the liquid or dried state.

B. During the survey, each sample will be analyzed for *Salmonella* spp., Enterobacteriaceae, generic *Escherichia coli* (*E. coli*), coliforms, and aerobic plate counts (APC).

C. The RLEBS will provide FSIS and the regulated industry with the data to understand the prevalence and quantitative levels of selected foodborne pathogens and indicator microorganisms. The data will enable the Agency and industry to work toward reducing the risk of foodborne pathogens in egg products and improving standards for thermal processing.

III. REFERENCES

[FSIS Directive 7355.1, Revision 2](#), *Use of Sample Seals for Laboratory Samples and Other Applications*.

[FSIS Directive 10,210.1](#), *Unified Sampling Form*.

[FSIS Directive 10,230.5 \(Attachment 1\)](#), *Salmonella Analysis – Collecting Raw Meat and Poultry Samples*.

IV. REVIEW OF FSIS DIRECTIVES AND TRAINING MATERIALS

Upon receipt of this notice, IPP are to review:

1. FSIS Directive 7355.1, Revision 2, *Use of Sample Seals for Laboratory Samples and Other Applications*;
2. The instructions for completing FSIS Form 10,210-3 in FSIS Directive 10,210.1, *Unified Sampling Form*; and
3. FSIS Directive 10,230.5 (Attachment 1), *Salmonella Analysis – Collecting Raw Meat and Poultry Samples*, Section 4.a, *Aseptic Sampling Techniques – Putting on gloves*, and Section 8, *Sample Storage Prior to Shipment*.

V. AWARENESS MEETING WITH PLANT MANAGEMENT

A. Upon issuance of this notice, IPP are to schedule an awareness meeting with plant management at plants that produce pasteurized and heat treated egg products to inform them that the plant will be part of the nationwide FSIS RLEBS and explain the program. IPP are to inform plant management that the baseline survey will begin on March 12, 2012.

B. IPP are to notify plant management in advance of the next meeting that this notice will be discussed and that the plant can access the notice on the FSIS website. IPP are to review the following program points with plant management during the awareness meeting:

1. The purpose of the FSIS RLEBS is to collect data concerning the prevalence and levels of selected foodborne microorganisms and indicator microorganisms in unpasteurized liquid whole eggs, egg yolks, and egg whites without added ingredients prior to pasteurization or heat treatment. After FSIS has completed the RLEBS, the Agency will publish the results in an RLEBS official baseline report and post it on the FSIS Web site. The report will present a summary of baseline findings on a national basis.
2. IPP are to emphasize to plant management that the RLEBS is not a regulatory program. Therefore, individual sample results from this baseline testing will not be the basis for regulatory actions, and plants do not need to hold product subject to RLEBS sampling.

3. Plants included in the RLEBS must pasteurize raw liquid eggs or heat treat dried egg product. Egg product plants that only break shell eggs, and do not pasteurize or heat treat egg products, are not included in the RLEBS.
4. The raw liquid egg products identified to be sampled for this study are liquid whole eggs, liquid egg whites, and liquid egg yolks that do not contain any added ingredients. Samples are to be taken before any ingredients are added and before the raw liquid egg product has been subjected to any further processing procedure (e.g., desugaring, reverse osmosis, pasteurization).
5. IPP will collect samples from the production shift indicated on the FSIS sample request form. If a plant has two shifts, IPP will receive separate forms indicating the specific shifts that are to be sampled.
6. Three plant locations have been identified from which samples may be collected, depending upon equipment design and accessibility for sampling. The preferred location of sampling is Option 1, followed by Option 2, and then Option 3.
 - a. Option 1: Balance Tank: The sample is to be collected at the balance tank (Figure 1). The sample is to be taken before the addition of any ingredients and prior to pasteurization.
 - b. Option 2: Silo/Tank: If it is not possible to collect the sample at the balance tank, then the sample is to be collected directly from the storage silo/tank before the addition of any ingredients and prior to pasteurization (Figure 2).
 - c. Option 3: Collection Pot: If it is not possible to collect the sample at the balance tank or the silo/tank, then the sample is to be collected at the collection pot during the breaking process (Figure 3).
7. During the survey, the FSIS contract laboratory will analyze each sample for *Salmonella* spp., Enterobacteriaceae, generic *E. coli*, coliforms, and APC.
8. FSIS will not publish individual plant results.
9. FSIS will not post microbiological results in LEARN.
10. IPP will send all samples to the FSIS contract laboratory in Section X of this notice.
11. The Agency expects each plant to carry out any sampling described in its food safety system or as required by regulation in 9 CFR Part 590 for liquid egg products, regardless of whether FSIS collects an FSIS RLEBS sample.

C. IPP are to document the meeting in a memorandum of interview (MOI). At a minimum, the MOI is to include a list of participants in the meeting, and recount all matters discussed between FSIS IPP and the plant. The MOI is to be kept in the

inspection file, with a copy provided to plant management.

VI. SAMPLING FORMS - FSIS FORM 10,210-3

A. IPP are to use the following methodology for collecting samples and completing FSIS Form 10,210-3 as directed in this notice. IPP will receive the pre-printed FSIS Form 10,210-3 in the mail from Headquarters.

B. *Block 14 -- Project Number.* The applicable project ID code will be pre-printed in this block for the sample to be collected. The project ID codes differentiate between liquid whole eggs, liquid egg whites, and liquid egg yolks, as well as Shift 1 and Shift 2, and will display on the sampling form as follows:

Baseline – First Shift	Baseline – Second Shift
B51WHIT1 – Liquid Egg White	B51WHIT2 – Liquid Egg White
B51WHOL1 – Liquid Whole Egg	B51WHOL2 – Liquid Whole Egg
B51YOLK1 – Liquid Egg Yolk	B51YOLK2 – Liquid Egg Yolk

C. *Block 18 – Additional Information.* This block will contain information on the type of sample to be collected, and the shift during which IPP are to collect the sample. For example, if the sample is to be collected during the first shift, the sample form will read: “COLLECTION: Sample raw unpasteurized liquid egg whites during SHIFT 1.”

D. *Block 21 – Product Temperature.* IPP are to record the temperature of the product, if known. IPP are to record the temperature on the thermometer from the wall silo or storage tank. IPP are not to take the temperature of the product in the sample cup. If the sample is taken directly from the collection pot, IPP are to write “*Temperature Not Available*” in Block 21. IPP are to record the temperature in degrees Fahrenheit (°F).

E. *Block 28 Remarks.* This block will contain a list of questions designed to guide data analysis for this survey. FSIS has revised or removed some of the questions listed in FSIS Notice 52-11. The following questions will appear in Block 28: The information requested is essential for the correct analysis of baseline data. The italicized text under the questions provides guidance and clarification to IPP, but will not appear on the sample form. IPP are to provide responses to each question:

1. Check the production shift during which the sample was collected?

Shift 1 _____ Shift 2 _____

Identify the production shift during which the sample was collected. The shift checked should match the shift number indicated in block 14 and block 18 on the sample request form. By checking the appropriate shift, you are confirming that you have collected the samples from the shift specified on FSIS Form 10,210-3.

2. What is the age of the shell eggs? _____

Write in the age of the shell eggs broken, from which the sample was collected. Example: Less than 1 day, 1-2 days, 1 week. For product received in tankers, refer to the production date/time on the USDA or Import Certificate received with the shipment.

3. What is the source of the sample collected?

- a. Shell eggs broken at the plant prior to pasteurization ___%
- b. Domestic unpasteurized egg product received by the plant ___%
- c. Imported unpasteurized egg product received by the plant ___%

Circle the source from which the sample was collected. Write in the percentage of liquid eggs that came from a, b, or c. If the sample collected originated from two or more sources, write in the percentage from each source. The percentages should add up to 100%.

4. Where was the sample collected? Check one.

Balance tank _____ Silo/Tank _____ Collection pot _____

Identify where the sample was collected. If the sample cannot be collected at the balance tank, silo/tank, or collection pot, send an email to the "Liquid Egg Baseline" mailbox in Outlook immediately.

5. What is the estimated volume of the sampled lot? _____ pounds

Record the volume (in pounds), in the space provided, of the lot from which the sample was collected. If the sample must be collected on the day prior to pasteurization, please estimate the anticipated size of the lot.

6. Record the time of sample collection. _____

Record the time of day the sample was collected. For example, if the sample was collected at 9:30 AM, enter 0930 or if 6:00 PM, enter 1800 in the space provided.

7. How much time has elapsed between the time breaking started and the sample was collected? _____ hours

For tankers, refer to the production date/time on the USDA or Import certificate received with the shipment.

8. Record the approximate amount of time between sample collection and when processing begins. _____

Record the approximate amount of time that elapses from the time the sample is

collected and before any further processing of the product begins. For example, if the sample is selected at 8:00 am, and pasteurization begins at 8:30 am, record 30 minutes. If the product is to be dried, record the time between when the product is sampled, and when the desugaring or pasteurization process begins. For the purpose of this notice, processing is specifically defined to mean before any desugaring, reverse osmosis, or addition of ingredients begins on the lot of product to be sampled.

VII. SAMPLE SUPPLIES

A. The sample supplies for the RLEBS are provided by the Midwestern Laboratory (MWL) and are intended only for collecting samples for the RLEBS. Sampling supplies and sample forms will arrive in separate shipments. Each shipping container will include the following supplies, and will accompany each baseline sample request:

1. M-20 Shipper box
2. (2) pairs of sterile gloves
3. (1) 500 ml sterile plastic sample jar with screw-cap
4. (1) 1 gallon zipper lock bag
5. (1) 6" x 12" plastic sleeve for FSIS Form 10,210-3 sample form
6. (1) cold pack (heat exchanger)
7. (1) cardboard spacer (to separate sample from the cold pack)
8. (6) FSIS Laboratory Sample Container Seal (FSIS Form 7355-2A/2B)
9. (1) Contract carrier (i.e., FedEx) billable stamp
10. (1) Absorbent pad

B. IPP at each plant sampling egg products for this baseline survey will receive a supply of sterile stainless steel 6 oz ladles, approximately one time every three months, during the course of this survey. The number of ladles shipped to each plant will correspond to the number of survey sample requests that have been identified for the plant during each three month period.

NOTE: Photos of the sampling supplies can be found in Appendix 1, Figure 4, and Table 1.

C. In addition to the sampling supplies listed above, IPP are to assemble the following additional supplies that will not be supplied by the MWL:

1. Paper towels to remove excess egg from the exterior of the sample container, if the sample is spilled;

2. Utility cart or available work space for staging the sampling supplies at the sample collection location; and
3. Caddy or bag for use in transporting the sampling supplies to the sample collection location.

VIII. BASELINE SAMPLING PROCEDURES

A. IPP assigned to plants in the RLEBS are to follow the sample collection methods in this notice and disregard the sample collection instructions in FSIS Directive 10,210.1, except where the instruction for completing FSIS Form 10,210-3, Sample Request Form, is referenced.

1. IPP are to verify when the sample is to be collected, and collect the sample specified on FSIS Form 10,210-3 during the designated time period indicated in Block 4 or at the next opportunity during the designated shift indicated by the Project ID Code printed in Block 14.

Sample collection is to be performed within two weeks after the date indicated in Block 4 of FSIS Form 10,210-3.

2. If IPP do not have access to unpasteurized egg product without added ingredients, IPP are to mark code 53 (miscellaneous discard code) in Block 33 of FSIS Form 10,210-3, indicate the reason for discard, and return the sample form to the MWL immediately. IPP are to send an email to the “Liquid Egg Baseline” mailbox in Outlook stating that the sample form was returned, and requesting instructions for returning the sample supplies. The IPP are to include his or her name, contact phone number, plant name, and plant number in the email.
3. If a plant does not pasteurize raw liquid eggs, IPP are to mark code 60 (plant does not produce the requested products) in Block 33 of FSIS Form 10,210-3, and return the sample form to the MWL immediately. IPP are to send an email to the “Liquid Egg Baseline” mailbox in Outlook stating that the sample form was returned and requesting instructions for returning the sample supplies. IPP are to state “Product Not Produced” in the subject line of the email. IPP are to include his or her name, contact phone number, plant name, and plant number in the email.
4. If a plant produces the egg product requested, but the product is not available during the sample collection period, IPP are to mark code 72 (requested sample unavailable during sampling period) in Block 33 of FSIS Form 10,210-3 and return the sample form to the MWL immediately. IPP are to send an email to the “Liquid Egg Baseline” mailbox in Outlook stating that the sample form was returned, and requesting instructions for returning the requested supplies. IPP are to include his or her name, contact phone number, plant name, and plant number in the email.

5. IPP are to collect 400 ml (3 full ladles) of the raw liquid egg product (specified on the sampling form) in the 500 ml sample jar at the balance tank, silo/tank, or collection pot, as necessary, immediately before pasteurization (See Appendix 1, Figures 5a and 5b, Notes for photos of how much liquid egg product is to be collected).

B. Before IPP collect the sample, IPP are to:

1. Verify that the sample box contains all the required supplies, upon receipt;
2. Ensure that all supplies needed for sample collection are available;
3. Place the cold packs in the freezer at least one day prior to sample collection. Pre-chill the sample shipping container, whenever possible, prior to sample collection;
4. Review FSIS Directive 10,230.5 (Attachment 1), *Salmonella Analysis – Collecting Raw Meat and Poultry Samples*, and follow the instructions in Section 8, *Sample Storage Prior to Shipment*, and
5. Ensure that sample shipping containers are not stored near heaters or in areas exposed to excessive heat.

NOTE: The laboratory will discard samples that arrive at or above 50°F. It is critical to the success of the RLEBS that sample temperature be properly maintained at or below 40°F during collection and shipment.

6. Identify the location from which to collect the sample:
 - a. Preferred Collection Location: Option 1 - Balance tank (See Appendix 1, Figure 1)
 - b. Alternative Collection Locations: Option 2 - Storage silo/tank sampling port or Option 3 - Collection pot. (See Appendix 1, Figures 2 and 3)
7. Verify that the appropriate Project ID Code and product type indicated on FSIS Form 10,210-3 corresponds with the type of egg product sample to be collected.
8. Verify the shift during which to collect the sample, as indicated in block 14 on FSIS Form 10,210-3.

C. When collecting the sample, IPP are to:

1. Collect samples during the first shift on Monday through Friday, if collected and shipped on the same calendar day;
2. Collect samples during the second shift on Monday through Thursday only; and

3. Collect sample before the addition of any ingredients and prior to the pasteurization process. Samples may be collected on the day prior to pasteurization when the plant's production practices or further processing procedures are such that a sample cannot be collected on the same day the product is to be pasteurized or heat treated (e.g., plant adds ingredients the day prior to pasteurization, product is subjected to reverse osmosis and then dried, or product is desugared and then dried).

NOTE: First-shift samples that are not collected and shipped on the same calendar day will be discarded by the laboratory. Second-shift samples that are collected and shipped the next calendar day will not be discarded by the laboratory.

D. When collecting the sample from the balance tank (Option 1), IPP are to:

1. Take the sampling supplies in a closed container or bag to the location where the sample will be collected.

IPP may need to use a utility cart or move a small table that can be sanitized to the sampling location so they have a flat work surface. IPP may also need to request the assistance of a Plant Quality Assurance person to assist them with sample collection.

2. Collect the sample after the balance tank is at least one-half full, or when the collection pot is full. If collecting a sample from a silo/tank, collect the product two hours after egg breaking has started. This is to ensure that the plant has accumulated enough liquid egg products for a sample to be collected.
3. Sanitize the work area, utility cart, or table top. Open the container used to transport the sampling supplies to the sampling location and take out the necessary sampling materials, making sure not to contaminate any of the materials.
4. Wash and dry hands thoroughly.
5. Carefully open the outer wrapper of the pouch containing the sterile stainless steel ladle so that the ladle can be easily lifted out once your hands are gloved. Do not touch or remove the sterile stainless steel ladle with your bare hands.
6. Carefully unscrew the lid of the sampling jar to loosen it, but do not remove it.

NOTE: The sampling jar will not be in a sterile pouch, but the sampling jar has been autoclaved. The inside of the jar is sterile.

7. Put on the sterile gloves, following the aseptic technique described in [FSIS Directive 10,230.5 \(Attachment 1\)](#), Section 4.a.

8. Remove the unscrewed sample jar lid, being careful not to touch the opening or the inside of the sample jar. Do not touch the inside of the sample jar lid. Set the jar lid on the sanitized work surface, face up, so that the inside of the lid is not touching the table.
9. Fill the sample jar with the same type of product as indicated on FSIS Form 10,210-3.
10. Dip the ladle into the balance tank and carefully pour the product into the sample jar until you have collected 400 ml (approximately 3 full ladles) (See Appendix 1, Figure 5a and 5b). Do not fill the sample jar over the balance tank or collection pot. Be careful not to spill product on the outside of the sample jar. Do not overfill the sample jar.

NOTE: If less than 400 ml of product is collected in the sample jar, the sample will not be analyzed.

11. After the sample has been collected, tightly screw the lid on the sample jar to ensure that the sample does not spill when transported. Ensure that the lid is threaded properly on the sample jar to prevent leakage of the sample. If the sample jar leaks during shipping, the laboratory will discard the sample. Wipe the outside of the sample jar with clean paper towels, if needed, to remove any egg product that may have spilled.
12. Place the closed sample jar into the zipper lock bag provided, squeeze out the air, and zip the bag closed.

NOTE: The sterile sampling ladles are to be used for the collection of only one sample. IPP are to discard the ladles following sample collections. Do not reuse the ladles for additional sample collection. Do not send the used ladles back to the laboratory.

E. When collecting the sample from the storage silo or tank (Option 2), IPP are to:

1. Follow steps 1 through 9, and 11 through 12 in Section VIII.D.
2. Allow approximately 500 ml of product to flow through the valve before collecting the sample in the sample jar to flush out the valve. IPP are to collect the product flushed through the valve into a container (i.e., a discard container or bucket) and dispose of the collected flushed product in the designated inedible container.
3. Place the sample jar under the valve and fill until 400 ml are collected. (See Appendix 1, Figures 5a and 5b.)
4. Tightly screw the lid on the sample jar. Wipe the outside of the sample jar with clean paper towels, if needed, to remove any egg product that may have spilled. Ensure that the lid is threaded properly on the sample jar to prevent leakage of

the sample. If the sample jar leaks during shipping, the laboratory will discard the sample.

F. When collecting the sample from the Collection Pot (Option 3), IPP are to follow the same procedures used when collecting the sample from the Balance Tank (Option 1) (see Section VIII.D).

G. Following sample collection, IPP are to immediately refrigerate the sample. The collected sample is to be held under refrigeration at 40°F, or lower, until shipped. Do not freeze samples. Keep all samples secure.

IX. SAMPLE STORAGE PRIOR TO SHIPMENT

IPP are to place the collected samples in the refrigerator and store at 40°F, or lower, until they are shipped. IPP are not to freeze the samples. IPP are not to store sample boxes near heaters or areas exposed to excessive heat. IPP are to safeguard the security of samples during preparation, storing, packaging, and submission of samples for testing (see [FSIS Directive 7355.1](#), *Use of Sample Seals for Program Samples and Other Applications*).

X. SAMPLE PACKING AND SHIPPING

A. IPP are to enter the appropriate information in Blocks 19, 20, 21, 26, and 28 through 32 of FSIS Form 10,210-3, and verify that the information is complete.

B. IPP are to follow the instructions provided in FSIS Directive 7355.1, *Use of Sample Seals for Laboratory Samples and Other Application*, on the use of sample seals (FSIS Form 7355-2A/2B) to maintain sample security and identification. To secure the sample, IPP are to:

1. Affix one small bar-coded label to the top center of the completed sample form and place the sample form in the plastic form sleeve;
2. Affix one small bar-coded label to the sample collection jar containing the collected sample, place the sample collection jar into the zipper lock bag provided, squeeze the air out of the bag, and zip the bag closed; and
3. Affix the corresponding medium-sized bar-coded FSIS Laboratory Sample Identification Label (FSIS Form -2B) on the zipper lock bag containing the sample jar.

C. To pack the shipping container when one cold pack is received, IPP are to:

1. Remove the cold pack from the freezer, place the absorbent pad in the shipping container, and place the cold pack on one of the sides of the container.

2. Place the cardboard separator between the cold pack and the sample jar to prevent the sample from freezing.
3. Place the sample collection jar (in the zipper lock bag) upright inside the shipper next to the cardboard separator and the cold pack.
4. Place the foam plug on top of the sample jar and press down slightly to secure contents.
5. Place FSIS Form 10,210-3 in its plastic sleeve on top of the foam plug.
6. Enter the required information on FSIS Laboratory Sample Container Seal (FSIS Form 7355-2A), and apply the seal to the box, using the instructions provided in FSIS Directive 7355.1.
7. Verify that the laboratory address on the contract carrier (i.e., FedEx) billable stamp and in Block 9 of FSIS Form 10,210.3 is as follows:

Food Safety-Net Services Ltd.
258 W. Turbo Drive
San Antonio, TX 78216
Attn: Robert Levy

8. Complete the return address fields on the contract carrier billable stamp and apply it to the shipping container.
9. Call the contract carrier to schedule pick up of the sample.
10. Maintain the shipping container in the refrigerator, and under FSIS control until the sample is picked up by the contract carrier.

NOTE: IPP are to use only the supplies provided for the RLEBS. Additional cardboard separators and gel coolant packs may be included with the sample supplies depending upon the time of the year. Sample supplies that are not provided in the shipping container sent from the MWL for this baseline study should not be used. If supplies have not arrived or are not complete, IPP are to send a request for the needed supplies through the “Liquid Egg Baseline” mailbox in Outlook.

NOTE: Appendix 1, Table 1 provides instruction on packing the sample in the shipping container when one or two cold packs are received.

D. If at any time IPP need to return supplies, they are to send an email to the “Liquid Egg Baseline” mailbox in Outlook to request instructions for return of supplies or shippers. IPP are to include his or her name, contact phone number, plant number, and name in the email. IPP will be sent FedEx Ground labels to use when returning sampling supplies. Sample supplies are to be sent via ground shipping to:

USDA, FSIS Midwestern Laboratory
Bldg 105-D Federal Center
4300 Goodfellow Road
St. Louis, MO 63120
Telephone: (314) 263-2680

XI. RESULTS FROM RLEBS BASELINE SAMPLES

IPP will not receive laboratory test results for samples analyzed at the contract laboratory for the FSIS RLEBS. These non-regulatory sample results are not posted in LEARN.

XII. DATA ANALYSIS

At the end of the nationwide FSIS RLEBS, the Office of Policy and Program Development, Office of Public Health and Science, and the Office of Data Integration and Food Protection will coordinate in analyzing the data from the survey. The primary purpose of these analyses is to estimate the national prevalence and levels of bacteria of public health concern in liquid egg products before either pasteurization or heat treatment and drying. These analyses will be used to guide policies related to microbial profiles in these products. The analyses may also be used to guide Agency procedures related to further sampling of liquid egg products intended for pasteurization or heat treatment and drying.

XIII. ADDITIONAL SUPPLIES, ASSISTANCE, AND INFORMATION

Questions concerning the training materials or the sampling procedures for the FSIS RLEBS and requests for additional sampling supplies, may be sent via email to the "Liquid Egg Baseline" mailbox in Outlook.

Refer questions regarding this notice to the Risk and Innovations Management Division through askFSIS at <http://askfsis.custhelp.com> or by telephone at 1-800-233-3935 by pressing 1 and then 4.



Assistant Administrator
Office of Policy and Program Development

Appendix 1

Figure 1: Balance Tank (Option 1): The balance tank is the preferred location to collect a sample. Collect the sample at the balance tank prior to the addition of added ingredients and immediately before pasteurization.

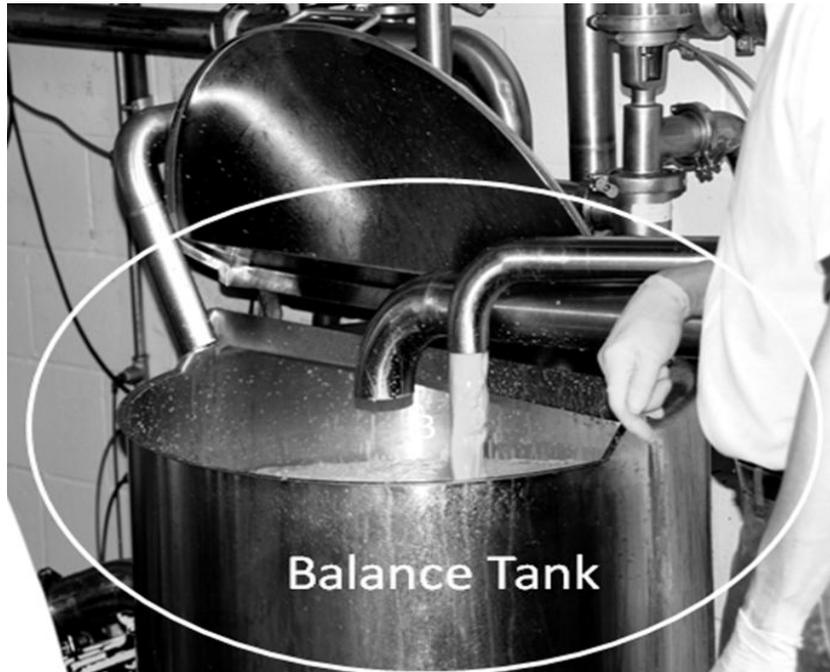


Figure 2: Raw Storage Silo/Tank (Option 2): The raw storage silo/tank is the second option for collecting a sample if the balance tank is not available. Collect the sample at the storage silo/tank before the addition of added ingredients and prior to pasteurization.



Figure 3: Collection Pot (Option 3): The collection pot should be sampled if IPP cannot collect the sample from the balance tank or the raw storage silo/tank. **This option is to be used only if the first two collection location options are completely unavailable.** Collect the sample at the collection pot during the breaking process and before the addition of added ingredients.



Figure 4: Sampling Supplies for the RLEB Survey: the bags, cold pack, FedEx billable stamp, FSIS sample seals, gloves, cardboard separator, sample jars, plastic form sleeve, and foam plug will arrive in the M-20 shipper sample box. The sterile ladles will be shipped in bulk, in the ladle box, separate from the M-20 shipper.



Figures 5a and 5b: Example of how to fill the sample jar: These photos show how to fill the sample jar. The jar is to be filled with approximately 3 full ladles (400 ml), to the level shown in Figure 5b.

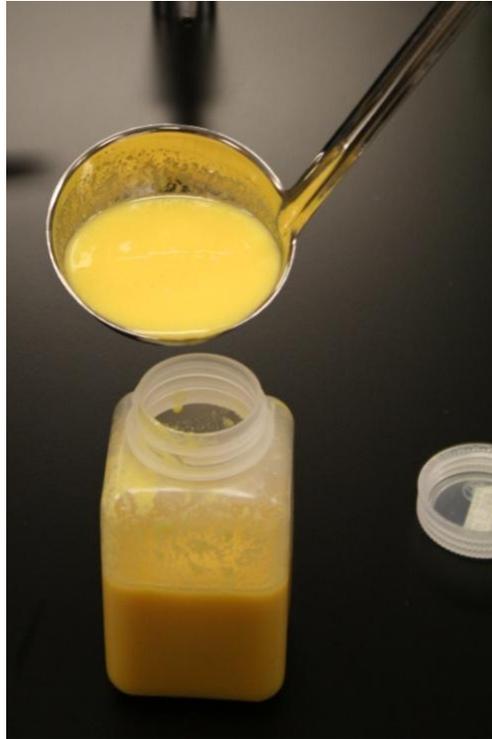


Figure 5a.



Figure 5b.

Table 1. Description of sample supplies

<p>The sample for the raw liquid egg baseline survey is collected in (1) sterile 500 ml sample jar. The sample jar must be filled with liquid whole egg, egg yolk or egg white without any added ingredients.</p> <p>The type of product will be specified on the FSIS sample form (FSIS Form 10,210-3).</p> <p>The shift during which the sample is to be collected will be specified on the sample form.</p>	 <p>FSIS Sample seals on appropriate items</p> <p>Full Sample Jar in Zipper Lock bag</p> <p>FedEx billable stamp</p> <p>Sample form in sample form sleeve</p>
<p>HOW TO PACK THE SHIPPING CONTAINER WHEN <u>ONE</u> COLD PACK IS RECEIVED: Place the absorbent pad in the bottom of the container, followed by cold pack, the cardboard separator, the sample jar (in the zipper lock bag), and then the foam plug. Place the completed sample form in its plastic sleeve on top of the foam plug. (Note: Do not return the ladle to the laboratory; it is to be discarded following sample collection).</p>	 <p>EXPRESS CARRIER BILL ONLY</p> <p>RLEBS</p>
<p>HOW TO PACK THE SHIPPING CONTAINER WHEN <u>TWO</u> COLD PACKS ARE RECEIVED: Place the absorbent pad in the bottom of the container, followed by a cold pack against the wall of the box, the cardboard separator, the sample jar (in the zipper lock bag), a cardboard separator, the cold pack on the opposite box wall, and place the foam plug on top. Place the completed sample form in its plastic sleeve on top of the foam plug. (Note: Do not return the ladle to the laboratory; it is to be discarded following sample collection).</p>	 <p>EXPRESS CARRIER BILL ONLY</p> <p>RLEBS</p>